

# LC-MS/MS Method for determination of capsaicin in edible oils and fats

Reference Standard: 《BJS 201801》

### **1** Preface

The spicy ingredient in peppers is mainly natural capsaicin, which is composed of 14 kinds of capsaicinoids. Among them, natural capsaicin and dihydrocapsaicin account for more than 90% of the total capsaicin. Synthetic capsaicin is a synthetic compound with biological functions similar to natural capsaicin. The active groups in the molecular structure are similar to natural capsaicin. However, in terms of spiciness and price, synthetic capsaicin is more popular, so there is a possibility of illegal use. Edible fats and oils do not come into contact with chili peppers during normal production and processing and do not contain capsaicin. The production level of recycled oil is not enough to completely remove capsaicin. Therefore, the quality of oil can be judged by measuring the capsaicin content in oil.

Based on "BJS 201801 Determination of Capsaicin in Edible Oils and Fats", this article established a rapid and highly sensitive detection method for the determination of three capsaicinoids in edible oils and fats using ultra-high performance liquid chromatography-triple quadrupole tandem mass spectrometer EXPEC 5210.

# 2 Experimental part

#### 2.1 Standards, reagents, and equipments

Standards: Synthetic capsaicin, natural capsaicin, and dihydrocapsaicin (Tianjin Alta Technology Co., Ltd., stored at  $-20^{\circ}$  C).

Reagents: Methanol, formic acid, acetonitrile, methylene chloride (chromatography grade), sodium hydroxide, concentrated sulfuric acid (analytical

grade), C18 solid phase extraction cartridge (500 mg, 6 mL).

Equipments: ULC 510 Ultra-high-performance liquid chromatograph (specifically equipped with a binary ultra-high-pressure infusion pump, ultra-highpressure autosampler, and column oven), and EXPEC 5210 LC-MSMS.



2.2	Liquid	phase	and	mass	spectro	metry	conditions
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	Table I Chiomatograp	ity and the MS co	Juditions				
	mobile phase	<ul><li>A: 0.1% Formic acid water</li><li>B: 0.1% Acetonitrile formate</li></ul>					
	flow speed		0.3 mL/min				
	column	C18 colum	n(2.1*100 mr	n, 1.8 µm)			
	Sample volumn		2 µL				
	Running time		7 min				
LC conditions		Time (min)	A (%)	B (%)			
		0	85	15			
		1	85	15			
	Elution gradient	2	10	90			
		5	10	90			
		5.1	85	15			
		7	85	15			
	Running mode		ESI+				
MS conditions	Capillary high pressure	4.8 kV					
	Desolvent gas flow		13 L/min				
	Anti-blowing flow		2 L/min				

ography and the MS conditions Table 1 Cl

Desolvent gas temperature	350 °С
Collision gas flow	0.5 mL/min
Atomizing gas flow	1.8 L/min

Multiple reaction monitoring (MRM), monitoring ion pair, collision voltage (CE) and other parameters for each compound are as follows:

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基本信息								
方法名称 Capsaicin BJS 201801								
修改原因								
了开碰撞气 □ MCA模式 智能MXM 添加片段 插入片段 删除片段	0	🗟 🔕 📴 🛛	🧟 🗒 循环的	时间 0.36				
方法片段信 开始时间(min) 结束时间(min) 离子化模式 扫描模式	一使	用调谐电压 扫描间	『鬲 0.005	s 数据类型	连续图 🗸	跨度 0		
1 0 6 ESI+ MEM		化合物名称	母离子质量	子离子质量	驻留时间(s)	锥孔电压	碰撞能量	分辨室
	1	Nonivamide	294.1	137.1	0.05	50	24	Unit-Unit
	2	Nonivamide	294.1	170.1	0.05	50	13	Unit-Unit
	3	Capsaicin	306.2	137.1	0.05	50	25	Unit-Unit
	<b>1</b> 4	Capsaicin	306.2	182.1	0.05	50	15	Unit-Unit
	5	Dihydrocapsa	308.1	137.1	0.05	50	19	Unit-Unit
			wards as	Constant of the	Contraction (		1 100	

Figure 1 Capsaicin monitoring ion pair and other mass spectrometry parameters

#### 2.3 Sample pretreatment

Weigh 1 g of soybean oil sample into a 15 mL stoppered plastic test tube, add 1 mL of methylene chloride and 3 mL of 2% sodium hydroxide solution, vortex for 10 min, centrifuge at 4000 r/min for 10 min, and take the upper aqueous phase; The residual organic phase was extracted twice with 2 mL of 2% sodium hydroxide solution. After combining the aqueous phases, adjust the pH to 2-3 with dilute sulfuric acid solution. Rinse the C18 SPE cartridge three times with 3 mL acetonitrile, and then rinse it twice with 3 mL pure water. Add the extract solution to the cartridge, 1-2 drops per second. After all the solution passes through the SPE cartridge, add 2 mL Elute with acetonitrile three times with 3 mL acetonitrile, and then rinse it twice with 3 mL extract solution to the cartridge, add 2 mL flute with a mL pure water. Add the extract solution to the cartridge, add 2 mL flute with 3 mL pure water. Add the extract solution to the cartridge, 1-2 drops per second. After all the solution to the cartridge, 1-2 drops per second. After all the extract solution to the cartridge, 1-2 drops per second. After all the extract solution to the cartridge, 1-2 drops per second. After all the extract solution to the cartridge, 1-2 drops per second. After all the solution passes through the SPE cartridge, add 2 mL Elute with 3 mL pure water. Add the extract solution to the cartridge, 1-2 drops per second. After all the solution passes through the SPE cartridge, add 2 mL Elute with acetonitrile three times, blow to near dryness with nitrogen at 50 ° C, 1 mL of methanol was dissolved and passed through a 0.22 m organic microporous filter, and placed in a sampling bottle for LC-MS/MS measurement.

# **3** Result

#### 3.1 Linearity

Accurately pipette appropriate amounts of capsaicin mixed standard stock solution into volumetric bottles, Use methanol to dilute it into a standard series of mixed working solutions with contents of 0.1, 0.2, 0.5, 1, 5, 10, 50, and 100 ng/mL. Inject samples according to the above method, take the quantitative ion chromatography peak area of each target substance as the ordinate (Y), and the mass concentration of the target standard solution as the abscissa (X). The standard curve fitted by the external standard method is shown in the figure below: the linear correlation coefficients  $R^2$  within the corresponding linear range are all above 0.995, indicating good linearity.



Figure 2 Capsaicin standard curve

#### **3.2 Sensitivity**

Determine the linear lowest point, follow S/N=3 and S/N=10 to calculate the detection limit and quantitation limit. The results are shown in Table 2 below: the detection limit of capsaicin is 0.012-0.015  $\mu$ g/kg; the quantitation limit is 0.039-0.050  $\mu$ g/kg.

	-	=	
	RT		
compound	(mi	LOD	LOQ
	n)		
Synthetic	3.5	0.013	0.042

Table 2 Detection limit and quantitation limit of capsaicin  $(0.1 \,\mu g/kg)$ 

capsaicin	5	SD-0.1 ESI+ TIC CV:50V CE:24V	合成铼翃素 MRM:294.1->137.1 1.85e4 cps 6.01 min 信号 18180 · · · · · · · · · · · · · · · · · · ·
Natural capsaicin	3.5 4	0.015 SD-0.1 ESI+ TIC CV:50V CE:25V	0.050   天然辣椒素 MRM:306.2->137.1 3.60e3 cps 6.01 min   3.60e3 cps 6.01 min   1.09   5.32   5.99   1.09   6 min   1 min
Dihydro capsaicin	3.6	0.012 SD-0.1 ESI+ TIC CV:50V CE:19V 100 80 80 80 40 20 0 0 1 2 3.64	O.039  二氢铼锶素 MRM:308.1->137.1 3.60e3 cps 6.01 min 信 号 5580 噪 声 100 信噪比发型 信噪比类型 100

# **3.3 Precision**

Configure a mixed standard solution of 0.5 ng/mL and 10 ng/mL capsaicin, and inject the sample 6 times continuously to check the repeatability of retention time and peak area. The results are as shown in the figure below: the RSD of the retention time is less than 0.1%, and the RSD of the peak area is less than 4.9 %, the instrument precision is good.

	Ľ.,		w.	样品信息	3	天然	陳椒素	二氢	谏椒素	合成報	刺素
	1	۴		数据文件	定量方法	保留时间	峰面积	保留时间	峰面积	保留时间	峰面积
1		٣		SD-0.5-1	辣椒素定量方法	3.54	20177.13	3.65	25386.90	3.55	6299.62
2		٣		SD-0.5-2	辣椒素定量方法	3.55	22387.53	3.65	25150.45	3.56	6655.44
3		٣		SD-0.5-3	辣椒素定量方法	3.55	20554.26	3.65	26344.27	3.56	6386.74
4		٣		SD-0.5-4	辣椒素定量方法	3.55	21576.67	3.65	26106.67	3.56	6809.48
5		٣		SD-0.5-5	辣椒素定量方法	3.55	21280.27	3.65	26378.01	3.55	6706.63
6		٣		SD-0.5-6	辣椒素定量方法	3.55	19561.20	3.65	23614.47	3.56	6484.20
Min				1		3.54	19561.20	3.65	23614.47	3.55	6299.62
Max						3.55	22387.53	3.65	26378.01	3.56	6809.48
AVG						3.55	20922.84	3.65	25496.79	3.56	6557.02
SD						0.00	1024.36	0.00	1052.79	0.00	198.17
• RSD						0.07	4.90	0.08	4.13	0.09	3.02
IDL						0.00	0.00	0.00	0.00	0.00	0.00

				样品信息	3	天然	辣椒素	二氢	辣椒素	合成辣椒素		
	1	🌹 🚩 数据文件 定里方法				保留时间	峰面积	保留时间	峰面积	保留时间	峰面积	
1		٣	V	SD-10-1	辣椒素定量方法	3.55	338876.80	3.65	402801.20	3.55	93797.98	
2		٣	1	SD-10-2	辣椒素定量方法	3.54	344471.20	3.65	409204.90	3.56	95098.86	
3		٣	1	SD-10-3	辣椒素定量方法	3.54	360624.80	3.64	426327.10	3.55	98825.51	
4		٣	Y SD-10-4		辣椒素定量方法	3. <mark>5</mark> 4	370649.30	3.65	444893.80	3.55	103886.10	
5		٣	V	SD-10-5	辣椒素定量方法	3.55	336166.30	3.65	411897.20	3.56	96002.66	
6		۴	7	SD-10-6	辣椒素定量方法	3.54	376479.60	3.65	446509.90	3.55	101897.10	
Min						3.54	336166.30	3.64	402801.20	3.55	93797.98	
Max						3.55	376479.60	3.65	446509.90	3.56	103886.10	
AVG						3.54	354544.70	3.65	423605.70	3.55	98251.37	
SD						0.00	17100.81	0.00	18774.92	0.00	4004.97	
► RSD						0.08	4.82	0.10	4.43	0.08	4.08	
IDL						0.00	0.00	0.00	0.00	0.00	0.00	

Figure 3 Retention time and peak area stability of 0.5 and 10 ng/mL mixed standard solutions

# 3.4 Sample test

Take 1g soybean oil sample (n=3), pre-process according to section 2.3 and then run the test on the machine. The blank and sample test results are as shown in the figure below: within the corresponding retention time, none of the three capsaicinoids were detected.



Figure 4 TIC spectra of blank and soybean oil samples

					样品信息			天然	辣椒素			二氢	陳椒素		合成辣椒素			
		1	٣		数据文件	定量方法	保留时间	峰面积	计算浓度	回收率(%)	保留时间	峰面积	计算浓度	回收率(%)	保留时间	峰面积	计算浓度	回收率(%)
•	1		۲	Z	油脂样品-1	辣椒素定量方法	3.55				3.65				3.56			
	2		٣	3	油脂样品-2	辣椒素定量方法	3.55				3.65				3.56			
	3		٣		油脂样品-3	辣椒素定量方法	3.55				3.66				3.55	1		

Figure 5 Soybean oil sample batch processing



Figure 6 50 ng/mL soybean oil sample spiked MRM overlay spectrum

#### **3.5 Accuracy**

Select capsaicin compound standard solutions with spiked concentrations of 5 and 40 ng/mL, use the same sample processing method, and inject 4 consecutive injections to examine the recovery rate of each concentration. The results are shown in the figure below: the recovery rate is above 70%.

					样品信息	2		二氯剂			天然	東椒素	sh	合成辣椒素				
		1	٣		数据文件	定重方法	保留时间	峰面积	计算浓度	回收率(%)	保留时间	峰面积	计算浓度	回收率(%)	保留时间	峰面积	计算浓度	回收室(%)
•	1		٣	V	油脂HS5-1	辣椒素回收率定量	3.64	43243.25	3.67	73.33	3.54	48340.79	3.63	72.50	3.55	248404.00	3.75	75.05
	2		٣	1	油脂HS5-2	辣椒素回收率定量	3.64	43885.86	3.75	75.01	3.54	47729.60	3.57	71.48	3.55	233849.70	3.51	70.14
	3		٣	V	油脂HS5-4	辣椒素回收率定量	3.65	42206.74	3.53	70.61	3.54	47453.41	3.55	71.02	3.55	245632.30	3.71	74.12
	4		٣	V	油脂HS5-3	辣椒素回收率定量	3.65	42638.25	3.59	71.74	3.54	48190.93	3.61	72.25	3.55	249097.30	3.76	75.29
	5		٣	V	油脂HS40-1	辣椒素回收率定量	3.65	257119.30	31.69	79.21	3.54	364696.60	29.96	74.90	3.55	1709741.00	28.42	71.06
	6		٣		油脂HS40-2	辣椒素回收率定量	3.65	245291.50	30.14	75.34	3.54	351907.80	28.90	72.24	3.55	1723902.00	28.66	71.66
	7		۲	V	油脂HS40-4	辣椒素回收率定量	3.65	248741.90	30.59	76.47	3.54	349071.50	28.66	71.65	3.55	1722600.00	28.64	71.60
	8		٣	V	油脂HS40-3	辣椒素回收率定量	3.65	257729.50	31.77	79.41	3.54	351511.30	28.86	72.16	3.55	1707724.00	28.39	70.98

Figure 7 Edible fat spiked recovery rate batch processing

# **4** Conclusion

This article established a method for the determination of capsaicin compounds in edible oils using the EXPEC 5210 triple quadrupole liquid mass spectrometer of EXPEC Technology, to examine the linearity, sensitivity, precision and recovery rate of the method. The results show that: the standard curves of the three capsaicin compounds have good linearity in the range of 0.1-100 ng/mL, and the correlation coefficients R2 are all greater than 0.995; The detection limit is 0.012-0.015 g/kg, the quantification limit is 0.039-0.050 g/kg; the retention time RSD is within 0.1%, and the peak area RSD is within 4.9%; The standard recovery rate is above 70%; all meet the standard requirements. This method can perform rapid, accurate and sensitive quantitative analysis of capsaicin compounds in edible oils and fats.